CosmoPAGE Bis-tris Precast Gel
- Bis-tris gel, 15 wells –

12 %, Cat. No. NU001215
4-12 %, Cat. No. NU041215

Introduction
CosmoPAGE Bis-Tris Precast Gel is a high-performance and easy to use precast polyacrylamide gel for electrophoresis in Bis-Tris buffer system (MOPS or MES). The optimized gel formula allows CosmoPAGE Bis-Tris Precast Gel to show improved resolution, accurate results, and an extended shelf-life over conventional gel. CosmoPAGE Bis-Tris Precast Gels are available in gradient (4 to 12%) and fixed (12%) concentrations of polyacrylamide in 15-well formats.

Product Information

Product	Cat. No.	Gel % / # of well	QTY
CosmoPAGE Bis-tris Precast Gel	NU001215	12% / 15 wells	10 gels
CosmoPAGE Bis-tris Precast Gel	NU041215	4 - 12 % / 15 wells	10 gels

Storage
4 °C

Shelf Life
12 months

Features
- Compatible with Bio-Rad Mini-PROTEAN® CORE
- No combs and tape
- Sharp, straight bands
- Numbered well; loading volume up to 28 µl/wells

Procedures

Running Buffer
Dissolve CosmoPAGE MOPS Running Buffer in 1000 ml of deionized water to make the 1X MOPS running buffer. Or prepare 10X stock running buffer with the following recipe:
10X MOPS running buffer:
Tris base 60.6 g, MOPS 104.6 g, SDS 10.0 g, EDTA 3.0 g.
Deionized water to 1000 ml.
10X MES running buffer preparation:
Tris base 60.6 g, MES 97.6 g, SDS 10.0 g, EDTA 3.0 g.
Deionized water to 1000 ml
*Always use fresh 1X running buffer

Sample Preparation
Mix sample with 2X sample buffer. Heat the diluted samples at 90-95°C for 5 min.
Recipe of 2X sample buffer with reducing agents:
62.5 mM Tris-HCl pH 6.8, 2% SDS, 25% (v/v) glycerol, 0.01% bromophenol blue, and 100 mM DTT (or 5% β-mercaptoethanol) as reducing agent.

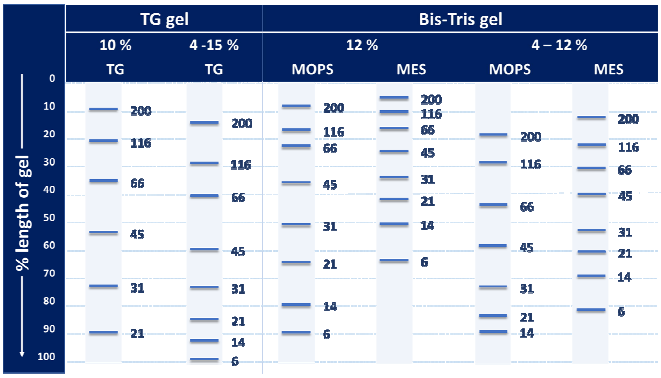
Setting CosmoPAGE Precast Gel into Bio-Rad Mini-PROTEAN® Core
Totally the same way to place CosmoPAGE precast gels and Bio-Rad Mini-PROTEAN gels; no adaptors required.

Running Conditions

	100 V	150 V	200 V
Running time	90 min.	60 min.	35 min.
Current			
Initial per gel	15-20 mA	50-60 mA	100-110 mA
Final per gel	5-10 mA	20-30 mA	30-40 mA
TEMP.	25°C	25-35°C	25-35°C

Running time might vary depending on power supply and the temperature of gel, running buffer and lab.

Gel Migration chart



Migration patterns of Unstained Protein Markers (#29458-24)

Gel staining
1. Insert a cassette opener into four corners of cassettes.
2. Carefully **pry but not twist** the opener to separate two plates.
3. Gently remove the gel from the plate.
4. Follow a procedure of most popular staining methods, such as Coomassie dyes (R-250 or G-250), Silver-stain solution, and fluorescent.

Gel blotting
Follow the general guidelines for your blotting apparatus; compatible to wet, semi-dry, or dry transferred to PVDF or nitrocellulose membrane.

Technical Support
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