CosmoPAGE TG Precast Gel
- Tris-glycine gel, 15 wells –

10 %, Cat. No. NU001015
4-15 %, Cat. No. NU041515

Introduction
CosmoPAGE TG Precast Gels (Tris-Glycine) are ready-to-use acrylamide gels for SDS-PAGE running in Tris-Glycine buffer system. With unique formula, CosmoPAGE TG Precast Gels perform enhanced resolution, sharper bands, and longer shelf life as compared with conventional Laemmli Tris-HCl gels. The protein migration patterns in CosmoPAGE TG series, however, are similar with typical Laemmli Tris-HCl gels, and thus CosmoPAGE TG Precast Gels are compatible to traditional SDS-PAGE and subsequent analyses.
CosmoPAGE TG Precast Gels are available in gradient (4 to 15%) and fixed (10%) concentrations of polyacrylamide in 15-well formats.

Product Information

Product	Cat. No.	Gel % / # of well	QTY
CosmoPAGE TG Precast Gel	NU001015	10 % / 15 wells	10 gels
CosmoPAGE TG Precast Gel	NU041515	4 - 15 % / 15 wells	10 gels

Storage
4 °C

Shelf Life
12 months

Features
- Compatible with Bio-Rad Mini-PROTEAN® CORE
- No combs and tape
- Sharp, straight bands
- Numbered well; loading volume up to 28 µl/wells

Procedures
Running Buffer
Prepare 10X stock running buffer with the following recipe:
Tris base 30.0 g, Glycine 144.0 g, SDS 10.0 g.
Deionized water to 1000 ml.
Dilute to 1X for use*
*Always use fresh 1X running

Sample Preparation
Mix sample with 2X sample buffer. Heat the diluted samples at 90-95°C for 5 min.
Recipe of 2X sample buffer with reducing agents:
62.5 mM Tris-HCl pH 6.8, 2% SDS, 25% (v/v) glycerol, 0.01% bromophenol blue, and 100 mM DTT (or 5% β-mercaptoethanol) as reducing agent.

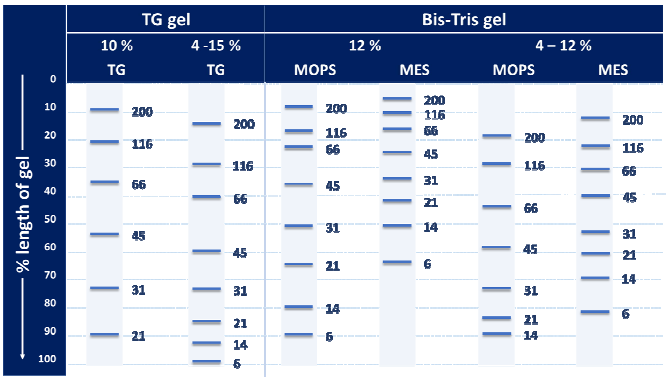
Setting CosmoPAGE Precast Gel into Bio-Rad Mini-PROTEAN® Core
Totally the same way to place CosmoPAGE precast gels and Bio-Rad Mini-PROTEAN gels; no adaptors required.

Running Conditions

	100 V	150 V	200 V
Running time	90 min.	60 min.	35 min.
Current			
Initial per gel	15-20 mA	50-60 mA	100-110 mA
Final per gel	5-10 mA	20-30 mA	30-40 mA
TEMP.	25°C	25-35°C	25-35°C

Running time might vary depending on power supply and the temperature of gel, running buffer and lab.

Gel Migration chart



Migration patterns of Unstained Protein Markers (#29458-24)

Gel staining
1. Insert a cassette opener into four corners of cassettes.
2. Carefully **pry but not twist** the opener to separate two plates.
3. Gently remove the gel from the plate.
4. Follow a procedure of most popular staining methods, such as Coomassie dyes (R-250 or G-250), Silver-stain solution, and fluorescent.

Gel blotting
Follow the general guidelines for your blotting apparatus; compatible to wet, semi-dry, or dry transferred to PVDF or nitrocellulose membrane.

Technical Support
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