

Protocol 014

Immobilization of Chemical Compounds on NHS Beads

Materials

1) Beads and Ligands

NHS beads (Product No.: TAS8848N1141): 10 mg

(Functional groups: 200–300 nmol/mg)

NHS beads are stored in IPA (isopropyl alcohol)

Chemical compounds: approximately 1 mg

*** Please keep in mind that the shelf life of NHS beads are <1 mo. Prolonged storage will result in loss of NHS esters impacting loading capacity.**

2) Reagents

N,N-Dimethylformamide (DMF)

2-Aminoethanol (ethanolamine) (M.W. 61.08)

Methanol

Ammonium acetate (M.W. 77.08)

Acetic acid (M.W. 60.05)

Acetonitrile

3) Apparatus

Micro centrifuge (HITACHI CF15RX2)

Microtube Mixer (TOMY MT-360)

Ultrasonic homogenizer (TAITEC VP-15 with cup horn)

HPLC system (Waters 2695 with 2998 PDA and SMH)

HPLC column (Waters Symmetry 5 μ m C18 4.6 \times 250mm WAT054275)

Methods

1) Immobilization of Chemical Compound on NHS Beads

Dissolve chemical compound in DMF

Prepare 5 mM chemical compound solution in DMF

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Start with 2.5 mg of NHS beads for each immobilization concentration

(NHS beads are suspended in IPA)

↓

Wash beads with DMF

(Centrifuge at 15,000 rpm for 5 min at R.T. and remove the supernatant)

↓

Suspend beads in each amount of DMF, and add each amount of 5 mM chemical compound solution as shown below

| Conc. of chemical compound at immobilization (mM) | 0 | 0.1 | 0.3 | 1 |
|---|-----|-----|-----|-----|
| DMF (μl) | 500 | 490 | 470 | 400 |
| 5 mM chemical compound solution (μl) | 0 | 10 | 30 | 100 |

↓

Mix for 70 min at R.T. by using Microtube Mixer

↓

Centrifuge at 15,000 rpm for 5 min at R.T.

↓

Transfer the supernatant (Sup A) to another tube for measurement of released NHS by HPLC [refer to Method 2) below for measurement instructions]

↓

Resuspend in 500 μl of 1 M ethanolamine in DMF (by using homogenizer)

↓

Mix for 2 h at R.T. by using Microtube Mixer

↓

Centrifuge at 15,000 rpm for 5min at R.T.

↓

Transfer the supernatant (Sup B) to another tube for measurement of released NHS by HPLC [refer to Method 2) below for measurement instructions]

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Wash with 500 μl of 50 % methanol in H₂O, 3 times

↓

Resuspend in 100 μ l of 50 % methanol in H₂O (by using homogenizer)

↓

Store at 4 °C

2) Determination of the Amount of Immobilized Chemical Compound

Prepare 10 mM ammonium acetate (pH 5.70)

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Prepare sample buffer and standard samples

Sample buffer: 47 ml of 10 mM ammonium acetate + 3 ml of acetonitrile (94 : 6)

Standard sample: 0, 10, 100, 1000, 5000 μ M NHS in DMF

↓

Mix 20 μ l of sample (standard sample, Sup A, or Sup B) and 180 μ l of sample buffer

↓

Filter at 5,000 \times g for 1 min by using 0.2 μ m centrifugal filter devices

↓

Place 150 μ l of the filtrate in autosampler of HPLC system

↓

Set HPLC conditions

- Gradient (40 min including column washing process):

| Time (min) | 10 mM ammonium acetate (%) | Acetonitrile (%) | Gradient curve |
|------------|----------------------------|------------------|----------------|
| 0 | 94 | 6 | - |
| 10 | 60 | 40 | Linear |
| 12 | 20 | 80 | Linear |
| 20 | 20 | 80 | Linear |
| 22 | 94 | 6 | Linear |
| 40 | 94 | 6 | Linear |

- Flow Rate: 1 ml/min
- Injection Volume: 50 μ l
- Temperature: 40 $^{\circ}$ C

↓

Run HPLC

↓

Analyze the amount of NHS in the samples from peak area at 260 nm

↓

Measure the amount of released NHS in Sup A and Sup B

↓

Amount of immobilized chemical compound is calculated from measurement of Sup A

(Amounts of carboxyl groups on COOH beads are calculated from measurement of Sup A and Sup B)